

# Mechanistic Insights into the H<sub>2</sub>S-Mediated Reduction of Aryl Azides Commonly Used in H<sub>2</sub>S Detection

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Supporting Information

ABSTRACT: Hydrogen sulfide (H2S) is an important biological mediator and has been at the center of a rapidly expanding field focused on understanding the biogenesis and action of H<sub>2</sub>S as well as other sulfur-related species. Concomitant with this expansion has been the development of new chemical tools for H2S research. The use of H2Sselective fluorescent probes that function by H2S-mediated reduction of fluorogenic aryl azides has emerged as one of the

most common methods for H<sub>2</sub>S detection. Despite this prevalence, the mechanism of this important reaction remains underscrutinized. Here we present a combined experimental and computational investigation of this mechanism. We establish that HS<sup>-</sup>, rather than diprotic H<sub>2</sub>S, is the active species required for aryl azide reduction. The hydrosulfide anion functions as a oneelectron reductant, resulting in the formation of polysulfide anions, such as HS2-, which were confirmed and trapped as organic polysulfides by benzyl chloride. The overall reaction is first-order in both azide and HS under the investigated experimental conditions with  $\Delta S^{\ddagger} = -14(2)$  eu and  $\Delta H^{\ddagger} = 13.8(5)$  kcal/mol in buffered aqueous solution. By using NBu<sub>4</sub>SH as the sulfide source, we were able to observe a reaction intermediate ( $\lambda_{max} = 473$  nm), which we attribute to formation of an anionic azidothiol intermediate. Our mechanistic investigations support that this intermediate is attacked by HS in the rate-limiting step of the reduction reaction. Complementing our experimental mechanistic investigations, we also performed DFT calculations at the B3LYP/6-31G(d,p), B3LYP/6-311++G(d,p), M06/TZVP, and M06/def2-TZVPD levels of theory applying the IEF-PCM water and MeCN solvation models, all of which support the experimentally determined reaction mechanism and provide cohesive mechanistic insights into H<sub>2</sub>S-mediated aryl azide reduction.

### INTRODUCTION

Hydrogen sulfide (H<sub>2</sub>S) has emerged as an important biological molecule of interest and joins nitric oxide (NO) and carbon monoxide (CO) as the third recognized gasotransmitter. Highlighting the importance of biological H<sub>2</sub>S, endogenous sulfide plays critical roles in the cardiovascular, gastrointestinal, immune, and nervous systems.<sup>2</sup> Enzymatic H<sub>2</sub>S synthesis occurs primarily from cystathionine- $\beta$ -synthase (CBS),<sup>3</sup> cystathionine-γ-lyase (CSE),<sup>4</sup> and 3-mercaptopyruvate sulfurtransferase (3-MST) working in concert with cysteine aminotransferase (CAT), although recent reports have also documented efficient H<sub>2</sub>S synthesis from D-Cys with D-amino acid oxidase (DAO) and 3-MST.<sup>5</sup> Because H<sub>2</sub>S is a weak acid, about 80% of H2S exists as monoanionic HS under physiological conditions. These different protonation states, along with the complex redox chemistry of sulfur, provide H<sub>2</sub>S with a more complex reactivity profile than that of NO or CO, resulting in reactions of H<sub>2</sub>S with both organic and inorganic targets. For example, the oxidative post-translational Spersulfidation of cysteine residues can enhance enzyme activity,6 and the reaction of H2S with metalloprotein transition-metal centers may also contribute to key signaling pathways. Although many of the molecular mechanisms of action remain to be elucidated, H2S is now known to be

involved in a diverse array of important biochemical processes including vasorelaxation, neuronal long-term potentiation, insulin regulation, angiogenesis, and inflammation. 10 Similarly, misregulation of endogenous H2S concentrations is associated with various conditions including hypertension, 11 Down syndrome, 12 and diabetes. 13

Paralleling the discovery of new biological roles of H<sub>2</sub>S has been the rapid expansion of chemical tools to study H<sub>2</sub>S in biological contexts. Such advances have included the development of chemical methods for H<sub>2</sub>S quantification, <sup>14</sup> methods of H<sub>2</sub>S delivery, <sup>15</sup> and reaction-based tools for H<sub>2</sub>S imaging. Fluorescent probe development has emerged as the most common imaging strategy and has relied on three main detection strategies, which include metal precipitation, 16 nucleophilic attack, 17 and reduction of azide or nitro groups. Of these methods, the H<sub>2</sub>S-mediated reduction of azides (Figure 1) has emerged as the most common strategy for H<sub>2</sub>S detection. In these systems, protection of a fluorogenic amine nitrogen as an azide renders the fluorophore nonemissive until the azide is reduced to the amine by H2S. Highlighting the prevalence and impact of this sensing strategy, over 65 papers

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# Azide-Based $H_2S$ Sensing $N_3$ $N_2$ $N_3$ $N_4$ $N_2$ $N_3$ $N_4$ $N_4$ $N_4$ $N_5$ $N_4$ $N_5$ $N_6$ $N_6$

# Model System for Mechanistic Studies

Figure 1. (top) Example of  $H_2S$ -mediated azide reduction with a fluorescent turn-on response. (bottom) Model azide system used here for mechanistic studies.

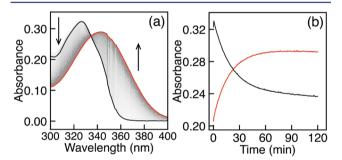
reporting azide-based  $H_2S$  probes have been published since the initial report of SF1 and SF2 in 2011. B Despite this widespread use, the mechanism by which  $H_2S$  reduces aryl azides remains underinvestigated, and key questions related to this process remain. For example, does  $H_2S$  act as a one- or two-electron reductant? Is  $H_2S(g)$  or  $HS^-$  the active sulfide species involved in the reduction? How many equivalents of  $H_2S$  are required for azide reduction? In addition to providing key insights into  $H_2S$  sensing, a functional understanding of how this reaction occurs would also provide valuable insights into the design of next-generation sensors. In view of this critical need, here we report experimental and computational investigations that provide new insights into the mechanism of  $H_2S$ -mediated azide reduction.

#### RESULTS AND DISCUSSION

On the basis of the high nucleophilicity of HS<sup>-</sup> as well as its prevalence under physiological conditions, it is likely that HSrather than H<sub>2</sub>S is the active species required for azide reduction. Drawing parallels to analogous reactions of azides with other nucleophilic reductants, such as phosphines in the Staudinger reaction, in which the terminal nitrogen is the site of nucleophilic attack, 19 our expectation is that the sulfidemediated reduction of aryl azides proceeds through initial nucleophilic attack on the electrophilic azide. After this initial attack to form an anionic azidothiol intermediate, we envisioned two possible mechanisms to generate the arylamine product (Scheme 1). The first mechanism proceeds through an intramolecular reaction in which proton transfer generates a terminal thiolate, which subsequently attacks the azide through either a four-membered triazathiocyclobutyl or three-membered diazathiocyclopropyl transition state to release N<sub>2</sub>, S<sup>0</sup>, and the arylamine product (mechanism I). In this scenario, H<sub>2</sub>S would act as a two-electron reductant, and a large, negative value of the entropy of activation ( $\Delta S^{\dagger}$ ) would be expected. By contrast to this intramolecular mechanism, the anionic azidothiol intermediate could be attacked by a second equivalent of HS-. In this scenario, protonation of the anionic azidothiol to generate a neutral azidothiol intermediate could enhance its electrophilicity, thus resulting in further activation for attack by the second equivalent of HS<sup>-</sup> to release N<sub>2</sub>, HS<sub>2</sub><sup>-</sup>, and the arylamine product (mechanism II). In this mechanism, H<sub>2</sub>S would act as a one-electron reductant, and a moderately

negative  $\Delta S^{\ddagger}$ , corresponding to a bimolecular reaction, would be expected.

Kinetic Studies of the  $H_2$ S-Mediated Reduction of C7-Az. To investigate the mechanism of sulfide-mediated azide reduction, we chose to use the previously reported 4-methyl-7-azidocoumarin (C7-Az) (Figure 1) as a model system because of its ease of synthesis and amenable spectroscopic properties. Treatment of aqueous buffered solutions of C7-Az with excess sodium hydrosulfide (NaSH) under pseudo-first-order conditions resulted in a decrease in the C7-Az absorbance ( $\lambda_{max} = 326$  nm) and concomitant formation of C7-NH<sub>2</sub> ( $\lambda_{max} = 343$  nm) with a well-anchored isosbestic point at 334 nm (Figure 2a). Under our experimental conditions, conversion to the

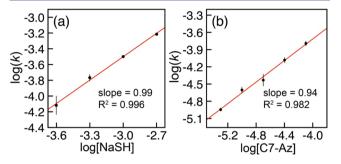


**Figure 2.** (a) UV–vis spectra and (b) time course data for the reaction of C7-Az (black) with NaSH to form C7-NH $_2$  (red) under pseudofirst-order conditions (20  $\mu$ M probe, 2.0 mM NaSH, 50 mM PIPES, 100 mM KCl, pH 7.4, 25 °C).

amine is complete within 60 min, making the reaction amenable for kinetic investigations. We also verified that the fluorescence turn-on profile of C7-Az matched the UV—vis data. Plots of the absorbances at 326 and 343 nm versus time (Figure 2b) both fit the first-order rate equation (Figure S1), and such fits were used to obtain pseudo first-order rate constants ( $k_{\rm obs}$ ) for subsequent experiments unless otherwise specified. Although sulfide-mediated azide reduction is likely a multistep process, no intermediates were observed by UV—vis spectroscopy.

**Reaction Order and Activation Parameters.** To determine the reaction orders with respect to [C7-Az] and [NaSH], we measured the rate of the reaction as a function of

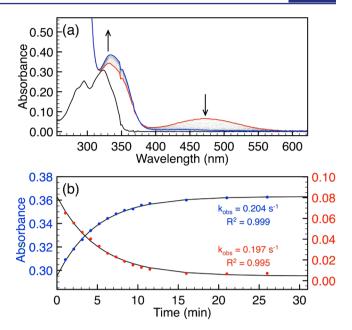
the reactant concentrations at constant pH and temperature. Working under pseudo first-order conditions in sulfide, varying [NaSH] allowed for determination of the [NaSH] order from a plot of  $\log(k_{\rm obs})$  versus  $\log({\rm [NaSH]})$ . This plot provided a linear relationship with a slope of 0.99, suggesting a first-order rate dependence on [NaSH] (Figures 3a and S3). To



**Figure 3.** (a) Plot of  $\log(k_{\rm obs})$  vs  $\log([{\rm NaSH}])$  for the reactions of 0.25–2.00 mM NaSH with 20  $\mu$ M C7-Az. (b) Plot of  $\log(k_{\rm obs})$  vs  $\log([{\rm C7-Az}])$  for the reactions of 2.00 mM NaSH with 5–80  $\mu$ M C7-Az. Standard conditions: 50 mM PIPES, 100 mM KCl, pH 7.4, 25 °C. Each measurement was repeated at least in triplicate.

determine the reaction order with respect to C7-Az, kinetic data were collected at constant pH, [NaSH], and temperature. To aid experimental measurement of  $k_{obs}$  for these experiments, we monitored initial rates because of the small changes in the absorbance of C7-Az and C7-NH2 during the course of the reaction. The resultant plot of  $log(k_{obs})$  versus log([C7-Az])also provided a linear plot with a slope of 0.94, which is also consistent with a first-order rate dependence on [C7-Az] (Figures 3b and S3). Supplementing the reaction order data, we also measured the rates of the reduction from 283-308 K under pseudo-first-order conditions in [NaSH] and calculated the second-order rate constants from the resultant  $k_{\rm obs}$  data, which ranged from 0.12 M<sup>-1</sup> s<sup>-1</sup> at 283 K to 0.95 M<sup>-1</sup> s<sup>-1</sup> at 308 K. Eyring analysis of the resultant second-order rate data revealed  $\Delta H^{\ddagger} = 13.8(5)$  kcal/mol and  $\Delta S^{\ddagger} = -14(2)$  eu (Figure S2).<sup>21</sup> The moderately negative  $\Delta S^{\ddagger}$  supports a ratelimiting step that proceeds through a bimolecular transition state rather than the highly strained intramolecular transition state associated with mechanism I.

Observation of Reaction Intermediates. To better isolate and differentiate the impacts of the H<sub>2</sub>S and HS<sup>-</sup> protonation states, we also investigated the reduction reaction in organic solution. Additionally, on the basis of the mechanisms proposed in Scheme 1, we envisioned that the use of an aprotic solvent could facilitate the observation of the proposed anionic intermediates. We chose to use MeCN as the solvent because of its high dielectric constant and aprotic nature. In support of our hypothesis that HS-, rather than  $H_2S(g)$ , is the active sulfide source in the reduction reaction, treatment of C7-Az with excess H<sub>2</sub>S(g) failed to produce any reaction. By contrast, addition of NBu<sub>4</sub>SH, which is a source of HS that is soluble in organic solvents, resulted in the immediate formation of an orange intermediate with an absorbance at 473 nm. During the course of the reaction, the absorbance of this highly colored intermediate slowly decayed to an absorbance corresponding to the C7-NH2 product (Figure 4a). A plot of the absorbance values for C7-NH<sub>2</sub> at 334 nm and the intermediate at 473 nm fit cleanly to a first-order rate equation with nearly identical  $k_{\rm obs}$  values of 0.204 and

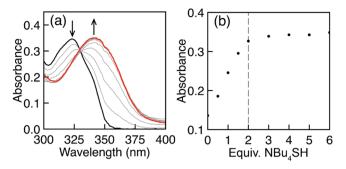


**Figure 4.** Reaction of C7-Az with excess NBu<sub>4</sub>SH (20  $\mu$ M C7-Az, 2.0 mM NBu<sub>4</sub>SH, MeCN): (a) full spectra overlay and (b) time courses at 334 (blue) and 473 (red) nm.

0.197 s<sup>-1</sup>, respectively (Figure 4b). These rate data support that rate-limiting decomposition of the observed intermediate leads to C7-NH<sub>2</sub> product formation. As control experiments, we confirmed that the absorbance at 473 nm does not correspond to other possible products formed during the course of the reaction, including hydropolysulfide anions (HS<sub>x</sub><sup>-</sup>), trisulfide radical ion (S<sub>3</sub><sup>•</sup>-), and deprotonated amine (C7-NH<sup>-</sup>) (Figures S5 and S7). In further support of the anionic structure of the intermediate, addition of an excess of H<sub>2</sub>O abolished the 473 nm absorbance and resulted in partial product formation, suggesting that protonation of the azidothiolate intermediate is required for subsequent sulfide attack (Figure S6). Using fluorescence spectroscopy, we also confirmed that the C7-Az fluorescence response is due to C7-NH<sub>2</sub> formation rather than formation of the intermediate.

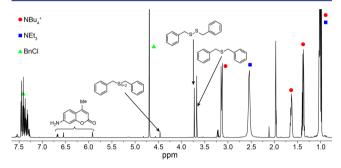
In addition to providing a solvent in which an intermediate could be observed, the use of aprotic MeCN in combination with NBu<sub>4</sub>SH allowed for titration of C7-Az to determine the reaction stoichiometry of HS $^-$  required to complete the reduction. These experiments demonstrated that complete conversion of the azide starting material to the amine product did not occur until 2 equiv of HS $^-$  had been added (Figure 5), which suggests not only that HS $^-$  functions as a one-electron reductant but also that 2 equiv of HS $^-$  is required for the reaction. The requirement of 2 equiv of HS $^-$  supports mechanism II (Scheme 1), in which HS $^-$  acts as a one electron reductant.

To gain further insights into the identity of the oxidized sulfur products formed during azide reduction, we treated C7-Az with 3 equiv of NBu<sub>4</sub>SH in MeCN and trapped the sulfur species present at the end of the reaction with benzyl chloride (BnCl). On the basis of the proposed reaction stoichiometry, we expected to observe 1 equiv of unreacted HS $^-$  and 1 equiv of oxidized sulfur products. Consistent with this hypothesis, we observed 1 equiv of Bn<sub>2</sub>S (3.67 ppm) and Bn<sub>2</sub>S<sub>2</sub> (3.72 ppm) as well as a small peak corresponding to higher-order benzyl polysulfides (4.45 ppm) (Figure 6). Importantly, the formation



**Figure 5.** (a) UV—vis titration showing the reduction of C7-Az (20  $\mu$ M in MeCN) to C7-NH<sub>2</sub> (red) by NBu<sub>4</sub>SH. (b) Monitoring the absorbance of the C7-NH<sub>2</sub> product as a function of NBu<sub>4</sub>SH establishes that 2 equiv of HS<sup>-</sup> are required for azide reduction.

of  $Bn_2S_2$  confirmed that  $HS_2^-$  was formed during the azide reduction reaction.



**Figure 6.** <sup>1</sup>H NMR spectrum (600 MHz) of the products formed during the reduction of C7-Az (12 mM) with NBu<sub>4</sub>SH (36 mM, 3 equiv) in CD<sub>3</sub>CN. Polysulfide anions formed during the reaction were trapped by addition of BnCl (12 equiv) and NEt<sub>3</sub> (12 equiv) after azide reduction was complete. The observation of Bn<sub>2</sub>S<sub>2</sub> and higher-order organic polysulfides supports the formation of HS<sub>x</sub><sup>-</sup> during the azide reduction process.

**Proposed Mechanism.** Taken together, the experimental results detailed above support mechanism II, which is outlined in greater detail in Scheme 2. The first step of the reaction proceeds by reversible nucleophilic attack of HS<sup>-</sup> on the terminal azide nitrogen of C7-Az (1), which is the most electrophilic nitrogen of the azide.<sup>22</sup> Nucleophilic attack on the terminal nitrogen has precedent in the Staudinger reaction<sup>19</sup> as well as the addition of carbon-based nucleophiles to azides.<sup>22</sup> This initial nucleophilic attack generates anionic azidothiol

intermediate 2, which we assign to the absorbance at 473 nm observed in MeCN on the basis of the experiments described above as well as qualitative time-dependent density functional theory (TD-DFT) calculations that support a bathochromic shift upon formation of the anionic azidothiol intermediate (Figure S9). Although anionic azidothiol 2 could be attacked directly by HS<sup>-</sup>, it is more plausible that 2 is transiently protonated to generate neutral 3, which would be significantly more electrophilic. Subsequent attack on 3 by a second equivalent of HS<sup>-</sup> then generates deprotonated amine 4, N<sub>2</sub>, and H<sub>2</sub>S<sub>2</sub>, after which proton transfer, likely mediated by the solvent, results in the formation of C7-NH<sub>2</sub> (5). We note that anion 4 is a significantly stronger base than intermediate 2 or HS<sub>2</sub><sup>-</sup> and should undergo facile proton transfer to generate 5. This proposed mechanism is consistent with the observed firstorder dependence of the reaction rate on HS<sup>-</sup> and azide as well as the moderately negative  $\Delta S^{\ddagger}$  for the reaction.

Rate Law Analysis. On the basis of the mechanism outlined in Scheme 2, the overall rate law for product formation can be derived from the reaction scheme shown in eq 1, in which 2 equiv of HS<sup>-</sup> and 1 equiv of aryl azide are involved in the reaction:

$$1 + HS^{-} \underset{k_{-1}}{\overset{k_1}{\rightleftharpoons}} 2 \underset{k_{-2}}{\overset{k_2[H^+]}{\rightleftharpoons}} 3 \xrightarrow{k_3[HS^-]} \text{product}$$
(1)

To simplify this reaction scheme, we note that conversion of 1 to 3 requires the net addition of  $H_2S(g)$  to 1, thus allowing for simplification of eq 1 to obtain eq 2:

$$1 + H_2S \underset{k'_{-1}}{\overset{k'_1}{\rightleftharpoons}} 3 \xrightarrow{k_3[HS^-]} \text{product}$$
(2)

Under this scenario, anionic azidothiol intermediate 3 is formed through a rapid pre-equilibrium before the rate-limiting attack by HS<sup>-</sup>, thus leading to the simplified steady-state rate law shown in eq 3:

rate = 
$$k_3[HS^-] \left( \frac{k_1'[ArN_3][H_2S]}{k_3[HS^-] + k_{-1}'} \right)$$
 (3)

In the limiting case in which  $k_3[HS^-] \ll k'_{-1}$ , which would result at low sulfide concentrations or if the unimolecular disassociation of azidothiol intermediate 3 is very fast, the rate law would condense to the form shown in eq 4:

rate = 
$$\frac{k'_1}{k'_{-1}} k_3 [ArN_3] [HS^-] [H_2S]$$
 (4)

#### Scheme 2

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Figure 7. Calculated potential energy surfaces for the proposed mechanism at the B3LYP/6-311++G(d,p) (black) and M06/def2-TZVPD (red) levels of theory with PCM solvation (H<sub>2</sub>O). Energies are reported in kcal/mol.

However, under conditions in which  $k_3[HS^-] \gg k'_{-1}$ , which would occur at higher [HS<sup>-</sup>], the rate law condenses to the form shown in eq 5:

$$rate = k_1'[ArN_3][H_2S]$$
 (5)

Under the experimental reaction conditions, we observe a first-order dependence of the rate on both [ArN<sub>3</sub>] and [HS<sup>-</sup>], which would be expected when [HS<sup>-</sup>] is in excess and is consistent with eq 5. Our attempts to shift the reaction to the other regime, in which a second-order dependence on sulfide is observed, including the use of C7-Az as the flooding substrate, maintained a first-order sulfide dependence, suggesting that we are experimentally unable to access the other limiting condition. However, when taken in combination with the reaction stoichiometry data and trapping of the  ${\rm HS}_2^-$  reaction product (vide supra) and DFT reaction coordinate calculations (vide infra), these kinetic data support the proposed mechanism for sulfide-mediated aryl azide reduction.

**DFT Calculations.** To provide added insights into the feasibility and energetics of the mechanism proposed in Scheme 2, we also conducted a full conformational search of each reactant, product, and intermediate and located each transition state using Gaussian 09 at the B3LYP/6-31G(d,p), B3LYP/6-311++G(d,p), M06/TZVP, and M06/def2-TZVPD levels of theory applying the IEF-PCM water and MeCN solvation models. The calculated potential energy surfaces (PESs) obtained at the B3LYP/6-311++G(d,p) and M06/def2-TZVPD levels of theory with the  $\rm H_2O$  solvation model are shown in Figure 7. Starting from C7-Az (1), the transition state of the first nucleophilic attack (TS<sub>12</sub>) was located with a calculated energy barrier of 9.1 kcal/mol, and from TS<sub>12</sub> the reaction proceeds to generate anionic azidothiol intermediate 2

with an overall endothermicity of 4.4 kcal/mol. Attempts to locate transition state structures derived from the attack of HSon anionic 2 were unsuccessful, whereas protonation of 2 to form neutral azidothiol intermediate 3 proceeds with a modest enthalpic penalty of 2.6 kcal/mol.<sup>23</sup> Subsequent nucleophilic attack by HS on 3 proceeds through transition state TS<sub>34</sub>, which has a local energy barrier of 9.2 kcal/mol to generate the deprotonated amine which is then protonated to generate the highly thermodynamically favored C7-NH<sub>2</sub> product (-59.5 kcal/mol). This second nucleophilic attack proceeds with the HS attacking the electrophilic sulfhydryl group of azidothiol intermediate 3 and is also consistent with the high homophilicity of sulfur. The calculated PES indicates that the second nucleophilic attack is the rate-determining step with an overall activation enthalpy of 16.2 kcal/mol, which is in good agreement with the experimentally measured  $\Delta H^{\ddagger}$  of 13.8(5) kcal/mol. No significant difference in the relative energies was observed when comparing the results between the DFT functionals and basis sets, highlighting the robust energy landscape derived from these computation studies. Although one would expect diffuse functions and dispersion forces to play an important role in this transformation, they have little impact on the thermochemistry of this PES (Figure S8). Significant efforts to locate transition state structures corresponding to the intramolecular 3- or 4-membered transitions states required in mechanism I were unsuccessful.

**Mechanistic Insights.** The experimental and computational investigations outlined above provide a cohesive mechanism for sulfide-mediated reduction of aryl azides. A number of key points are highlighted below. First, the mechanistic studies establish that  $HS^-$ , rather than  $H_2S$ , is the active species responsible for sulfide-mediated azide

reduction. Because HS is the dominant species under physiological conditions, these results highlight that the azide reduction method reports on the most prevalent form of available sulfide. Additionally, the initial addition of HS to the azide is reversible, suggesting that generation of the important azidothiol intermediate may be difficult under conditions where the sulfide concentrations are very low. It is possible that other sulfhydryl-containing biomolecules, such as glutathione, may participate in the reaction after the initial attack by the more nucleophilic hydrosulfide anion. Furthermore, the mechanistic insights suggest that protonation of the azidothiolate intermediate is an important step in the reaction. One would expect that addition of functional groups that could make this intermediate more basic and favor protonation would help to drive the reaction, thus facilitating nucleophilic attack in the subsequent and rate-limiting step. Additionally, these mechanistic studies show that HS<sup>-</sup> is consumed during each reduction of an aryl azide, highlighting that such sensing methods perturb the sulfide levels and the redox state of the system. On the basis of the growing importance of the reduction-labile sulfane sulfur pool, such reaction-based detection methods may inadvertently disturb the distribution of sulfide between the bound and free pools, thus opening new questions related to how H2S sensors impact redox homeostasis. With the absence of reversible H<sub>2</sub>S sensing mechanisms that operate under physiological conditions, such perturbations remain a reality of reaction-based sensors and need to be considered for biological inquiries in which irreversible reaction-based reporters are used.

#### CONCLUSIONS

Our mechanistic investigations into the sulfide-mediated reduction of aryl azides have revealed that this reaction is first-order with respect to C7-Az and NaSH under our experimental conditions and proceeds with activation parameters consistent with a biomolecular mechanism ( $\Delta S^{\dagger}$  = -14(2) eu) with a moderate enthalpic barrier ( $\Delta H^{\ddagger} = 13.8(5)$ kcal/mol). Reactions of C7-Az with NBu<sub>4</sub>SH in MeCN allowed for the observation of an absorbance that we attribute to the anionic azidothiol intermediate, and HS- titrations in this system established that 2 equiv of HS<sup>-</sup> is required for complete reduction, which is consistent with sulfide acting as a oneelectron reductant. Computational studies support the experimental observations and indicate that the second attack of sulfide is the rate-determining-step of the reduction, and the calculated activation energy (16.2 kcal/mol) is in good agreement with the experimental enthalpy of activation (13.8(5) kcal/mol). Taken together, the results of these mechanistic studies provide new insights into the individual steps required for H<sub>2</sub>S-mediated azide reduction and may be useful in the refinement of this important sensing strategy for H<sub>2</sub>S.

# **■ EXPERIMENTAL DETAILS**

Materials and Methods. All manipulations were performed under an inert atmosphere using standard Schlenk techniques or an Innovative Atmospheres N<sub>2</sub>-filled glovebox with O<sub>2</sub> levels less than 1.0 ppm. Anhydrous and air-free MeCN was obtained from a Pure Process Technologies solvent purification system. Deuterated solvents were dried by distillation over the appropriate drying agent and deoxygenated by three freeze-pump-thaw cycles. Tetrabutylammonium hydrosulfide (NBu<sub>4</sub>SH) and C7-Az were prepared as described in the literature.<sup>24</sup> Anhydrous sodium hydrogen sulfide (NaSH) was purchased from Strem Chemicals and handled under nitrogen. Piperazine-N,N'-bis(2-ethanesulfonic acid) (PIPES) and potassium

chloride (99.999%), both obtained from Aldrich, were used to make buffered solutions (50 mM PIPES, 100 mM KCl, pH 7.4) with Millipore water. Buffered solutions were degassed by vigorous sparging with  $\rm N_2$  and stored in an inert atmosphere glovebox. Stock solutions of C7-Az were prepared in MeCN or dimethyl sulfoxide. Gas-tight Hamilton syringes were used for solution transfers outside of the glovebox.

**Spectroscopic Methods.** UV—vis measurements were performed on an Agilent Cary 100 UV—vis spectrophotometer equipped with a QNW dual-cuvette temperature controller at  $25.00 \pm 0.05$  °C. All of the cuvette-based spectroscopic measurements were made under anaerobic conditions with solutions prepared under an inert atmosphere in 1.0 cm path length septum-sealed cuvettes obtained from Starna Scientific and were repeated at least in duplicate. NMR spectra were acquired on a Bruker Avance-III-HD 600 spectrometer with a Prodigy multinuclear broadband cryoprobe at 25.0 °C.

Computational Details. Calculations were performed using the Gaussian 09 software package. Chemcraft was used to analyze the results and generate graphical representations. Conformational searches were conducted at the B3LYP/6-311++G(d,p), M06/TZVP, and M06/def2-TZVPD levels of theory applying the IEF-PCM water and MeCN solvation models by exploring reasonable input dihedral angles. Frequency calculations were performed on each located stationary point to confirm that it was either a local minimum or a saddle point. Relative energies were calculated using the zero-point-energy-corrected electronic energies obtained in the frequency calculations.

General Conditions for UV–Vis Experiments. In a septum-sealed cuvette, 2.970 mL of a 20  $\mu$ M stock solution of C7-Az in either MeCN or buffered water was treated with 30  $\mu$ L of 200 mM HS<sup>-</sup> stock solution. The UV–vis spectra were monitored as a function of time, and the C7-NH<sub>2</sub> absorbance (343 nm) was used to measure the observed rate constants.

**General Conditions for NMR Experiments.** A septum-sealed NMR tube was charged with C7-Az (12 mM in 0.5 mL MeCN- $d_3$ ) under a nitrogen atmosphere, and 6.6  $\mu$ L of triethylamine was added. A 50  $\mu$ L aliquot of a MeCN- $d_3$  solution containing 240 mM NBu<sub>4</sub>SH was added. The azide was completely reduced upon sulfide addition, and BnCl (5.5  $\mu$ L) was added to trap the generated HS<sub>2</sub><sup>-</sup>. Authentic Bn<sub>2</sub>S<sub>2</sub> was subsequently added to confirm the identity of the singlet at 3.72 ppm. All additions were performed using Hamilton syringes.

#### ASSOCIATED CONTENT

# S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/jacs.5b10675.

UV-vis spectra, Eyring plot, control experiments, computational data, and a list of references using azide reduction for H<sub>2</sub>S sensing (PDF)

#### AUTHOR INFORMATION

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#### Note:

The authors declare no competing financial interest.

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# **■** REFERENCES

(1) Wang, R. Physiol. Rev. 2012, 92, 791-896.

- (2) We will use " $H_2S$ " to denote the totality of the sulfide pool, which includes  $H_2S$ ,  $HS^-$ , and  $S^{2-}$ . When we are clearly defining one protonation state, we will use  $H_2S(g)$  or  $HS^-$  for added clarity.
- (3) Abe, K.; Kimura, H. J. Neurosci. 1996, 16, 1066-1071.
- (4) Hosoki, R.; Matsuki, N.; Kimura, H. Biochem. Biophys. Res. Commun. 1997, 237, 527-531.
- (5) Shibuya, N.; Tanaka, M.; Yoshida, M.; Ogasawara, Y.; Togawa, T.; Ishii, K.; Kimura, H. *Antioxid. Redox Signaling* **2009**, *11*, 703–714.
- (6) Mustafa, A. K.; Gadalla, M. M.; Sen, N.; Kim, S.; Mu, W. T.; Gazi, S. K.; Barrow, R. K.; Yang, G. D.; Wang, R.; Snyder, S. H. Sci. Signaling **2009**, 2, ra72.
- (7) (a) Dorman, D. C.; Moulin, F. J.-M.; McManus, B. E.; Mahle, K. C.; James, R. A.; Struve, M. F. *Toxicol. Sci.* 2002, 65, 18–25.
  (b) Collman, J. P.; Ghosh, S.; Dey, A.; Decreau, R. A. *Proc. Natl. Acad. Sci. U. S. A.* 2009, 106, 22090–22095.
- (8) (a) Yang, G.; Yang, W.; Wu, L.; Wang, R. J. Biol. Chem. 2007, 282, 16567–16576. (b) Yang, W.; Yang, G. D.; Jia, X. M.; Wu, L. Y.; Wang, R. J. Physiol. 2005, 569, 519–531.
- (9) (a) Papapetropoulos, A.; Pyriochou, A.; Altaany, Z.; Yang, G. D.; Marazioti, A.; Zhou, Z. M.; Jeschke, M. G.; Branski, L. K.; Herndon, D. N.; Wang, R.; Szabo, C. *Proc. Natl. Acad. Sci. U. S. A.* **2009**, *106*, 21972–21977. (b) Cai, W. J.; Wang, M. J.; Moore, P. K.; Jin, H. M.; Yao, T.; Zhu, Y. C. *Cardiovasc. Res.* **2007**, *76*, 29–40.
- (10) Kaneko, Y.; Kimura, Y.; Kimura, H.; Niki, I. Diabetes 2006, 55, 1391-1397.
- (11) Yang, G.; Wu, L.; Jiang, B.; Yang, W.; Qi, J.; Cao, K.; Meng, Q.; Mustafa, A. K.; Mu, W.; Zhang, S.; Snyder, S. H.; Wang, R. *Science* **2008**, 322, 587–590.
- (12) Kamoun, P.; Belardinelli, M. C.; Chabli, A.; Lallouchi, K.; Chadefaux-Vekemans, B. Am. J. Med. Genet. Part A 2003, 116A, 310–311.
- (13) Wu, L. Y.; Yang, W.; Jia, X. M.; Yang, G. D.; Duridanova, D.; Cao, K.; Wang, R. Lab. Invest. **2009**, 89, 59–67.
- (14) (a) Wintner, E. A.; Deckwerth, T. L.; Langston, W.; Bengtsson, A.; Leviten, D.; Hill, P.; Insko, M. A.; Dumpit, R.; VandenEkart, E.; Toombs, C. F.; Szabo, C. Br. J. Pharmacol. 2010, 160, 941–957. (b) Shen, X.; Pattillo, C. B.; Pardue, S.; Bir, S. C.; Wang, R.; Kevil, C. G. Free Radical Biol. Med. 2011, 50, 1021–1031. (c) Shen, X.; Peter, E. A.; Bir, S.; Wang, R.; Kevil, C. G. Free Radical Biol. Med. 2012, 52, 2276–2283. (d) Montoya, L. A.; Pearce, T. F.; Hansen, R. J.; Zakharov, L. N.; Pluth, M. D. J. Org. Chem. 2013, 78, 6550–6557. (e) Montoya, L. A.; Shen, X.; McDermott, J. J.; Kevil, C. G.; Pluth, M. D. Chem. Sci. 2015, 6, 294–300.
- (15) (a) Benavides, G. A.; Squadrito, G. L.; Mills, R. W.; Patel, H. D.; Isbell, T. S.; Patel, R. P.; Darley-Usmar, V. M.; Doeller, J. E.; Kraus, D. W. Proc. Natl. Acad. Sci. U. S. A. 2007, 104, 17977–17982. (b) Song, Z. J.; Ng, M. Y.; Lee, Z.-W.; Dai, W.; Hagen, T.; Moore, P. K.; Huang, D.; Deng, L.-W.; Tan, C.-H. MedChemComm 2014, 5, 557–570. (c) Wallace, J. L.; Wang, R. Nat. Rev. Drug Discovery 2015, 14, 329–345. (d) Li, L.; Whiteman, M.; Guan, Y. Y.; Neo, K. L.; Cheng, Y.; Lee, S. W.; Zhao, Y.; Baskar, R.; Tan, C. H.; Moore, P. K. Circulation 2008, 117, 2351–2360. (e) Zhao, Y.; Wang, H.; Xian, M. J. Am. Chem. Soc. 2011, 133, 15–17. (f) Artaud, I.; Galardon, E. ChemBioChem 2014, 15, 2361–2364. (g) Le Trionnaire, S.; Perry, A.; Szczesny, B.; Szabo, C.; Winyard, P. G.; Whatmore, J. L.; Wood, M. E.; Whiteman, M. MedChemComm 2014, 5, 728–736.
- (16) Sasakura, K.; Hanaoka, K.; Shibuya, N.; Mikami, Y.; Kimura, Y.; Komatsu, T.; Ueno, T.; Terai, T.; Kimura, H.; Nagano, T. *J. Am. Chem. Soc.* **2011**, *133*, 18003–18005.
- (17) (a) Liu, C. R.; Pan, J.; Li, S.; Zhao, Y.; Wu, L. Y.; Berkman, C. E.; Whorton, A. R.; Xian, M. Angew. Chem., Int. Ed. 2011, 50, 10327—10329. (b) Qian, Y.; Karpus, J.; Kabil, O.; Zhang, S.-Y.; Zhu, H.-L.; Banerjee, R.; Zhao, J.; He, C. Nat. Commun. 2011, 2, 495. (c) Chen, Y. C.; Zhu, C. C.; Yang, Z. H.; Chen, J. J.; He, Y. F.; Jiao, Y.; He, W. J.; Qiu, L.; Cen, J. J.; Guo, Z. J. Angew. Chem., Int. Ed. 2013, 52, 1688—1691.
- (18) See the Supporting Information for a list of citations using  $H_2S$ -mediated azide reduction as a sensing strategy.

- (19) (a) Gololobov, Y. G.; Zhmurova, I. N.; Kasukhin, L. F. Tetrahedron 1981, 37, 437–472. (b) Gololobov, Y. G.; Kasukhin, L. F. Tetrahedron 1992, 48, 1353–1406. (c) Leffler, J. E.; Temple, R. D. J. Am. Chem. Soc. 1967, 89, 5235–5246. (d) Leffler, J. E.; Tsuno, Y. J. Org. Chem. 1963, 28, 902–906. (e) Fortman, G. C.; Captain, B.; Hoff, C. D. Inorg. Chem. 2009, 48, 1808–1810.
- (20) Chen, B.; Li, W.; Lv, C.; Zhao, M.; Jin, H.; Jin, H.; Du, J.; Zhang, L.; Tang, X. *Analyst* **2013**, *138*, 946–951.
- (21) Eyring analysis of the pseudo-first-order rate data afforded  $\Delta H^{\ddagger}$  = 13.8(5) kcal/mol and  $\Delta S^{\ddagger}$  = -26(2) eu, which is consistent with the change in standard-state entropy.
- (22) Brase, S.; Gil, C.; Knepper, K.; Zimmermann, V. Angew. Chem., Int. Ed. 2005, 44, 5188–5240.
- (23) For simplicity and to maintain atomic balance in the PES, we assumed that this protonation proceeded from  $H_2S$ .
- (24) (a) Thorson, M. K.; Majtan, T.; Kraus, J. P.; Barrios, A. M. Angew. Chem., Int. Ed. 2013, 52, 4641–4644. (b) Hartle, M. D.; Meininger, D. J.; Zakharov, L. N.; Tonzetich, Z. J.; Pluth, M. D. Dalton. Trans. 2015, 44, 19782–19785.
- (25) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, I.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, J. M.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth, G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian 09, revision C.01; Gaussian, Inc.: Wallingford, CT, 2009.